

CARBON-13 NUCLEAR MAGNETIC RESONANCE SPECTRA OF CARDENOLIDES

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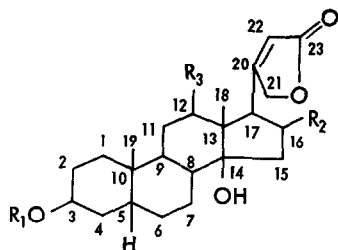
It has been demonstrated recently that natural-abundance  $^{13}\text{C}$  NMR spectroscopy is extremely useful for investigating stereochemical and mechanistic problems in steroid chemistry (1). Although several papers have presented correlations of structure with  $^{13}\text{C}$  NMR spectra for steroids having various types of ring junctions, such studies have not yet been made for biologically important cardenolides (2). In this paper, we report a complete  $^{13}\text{C}$  NMR analysis (3) for the carbons of several cardenolides (I-X). Cardenolides all occur as plant glycosides, and therefore, the present  $^{13}\text{C}$  NMR spectroscopic data will be useful for determining their structure and sequence, and also those of related natural products, without hydrolytic or enzymatic cleavage of their sugar moieties (4).<sup>†</sup>

Application of chemical shift theory (5), single-frequency off-resonance decoupling (SFORD), and noise off-resonance decoupling, and comparison with the spectra of structurally related compounds led to the assignments shown in the TABLE.

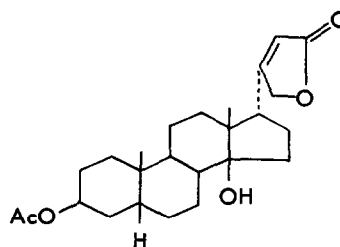
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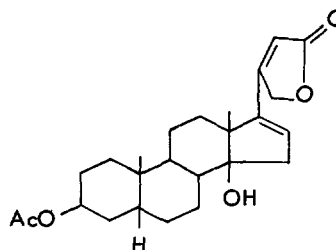
<sup>†</sup> Partially-relaxed-Fourier-transform experiments on some steroidal trisaccharides indicated that relaxation times in a mixed solvent of  $\text{CDCl}_3$  and  $\text{CD}_3\text{OD}$  (1.5 : 1) are markedly shorter than those reported for a disaccharide at comparable concentrations in an aqueous solution (6). Detailed results will be presented in a forthcoming paper.



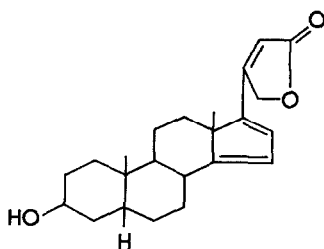
- I,  $R_1 = R_2 = R_3 = H$ : digitoxigenin  
 II,  $R_1 = R_3 = H$ ;  $R_2 = OH$ : gitoxigenin  
 III,  $R_1 = R_3 = H$ ;  $R_2 = OAc$   
 IV,  $R_1 = Ac$ ;  $R_2 = OAc$ ;  $R_3 = H$   
 V,  $R_1 = Ac$ ;  $R_2 = R_3 = H$   
 IX,  $R_1 = R_2 = H$ ;  $R_3 = OH$ : digoxigenin



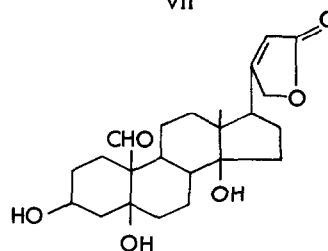
VI



VII



VIII



X: strophanthidin

Compared with *A/B-cis* *C/D-trans* steroids (1b, 1c), the major  $^{13}C$  NMR differences for the cardenolides are the expected shieldings for C-7, C-9, and C-16 and deshieldings for C-8, C-13, C-14, and C-15. The upfield shifts for the C-7 and C-9 signals are due to steric interaction between the axial hydrogens at these carbons and the C(14)-C(15) bond. Due to the equatorial  $\beta$ -effect of the C-14 $\beta$  hydroxyl group, the highest-field methine carbon signal does not arise from C-8 as in other steroids (1).

Chemical shift changes attendant with acetylation yielded the shift values for C-2, C-4, C-15, and C-17. Being unaffected by stereochemical changes, C-6, C-7, and C-11 varied little in their chemical shifts through most of the series. Of these signals, the C-6 signal is at the lowest field, whereas the C-7 and C-11 signals are undifferentiable in several compounds. While the axial hydrogens of C-11 and C-7 are

TABLE  
<sup>13</sup>C Chemical Shift Data on Cardenolides (δ in ppm downfield from TMS)

	I	II	III	IV	V	VI	VII	VIII	IX	X
C-1	30.0	30.0	30.0	30.7	30.8	30.8	30.8	30.7	30.0	24.8
C-2	28.0	28.0	27.9	25.2	25.4	25.3	25.4	27.9	27.9	27.4 <sup>a</sup>
C-3	66.8	66.8	66.8	71.1	71.4	71.3	71.3	66.7	66.6	67.2
C-4	33.5	33.5	33.4	30.7	30.8	30.8	30.8	33.5	33.3	38.1
C-5	35.9 <sup>a</sup>	36.4	36.4	37.2	37.4	37.4	37.3	36.8 <sup>a</sup>	36.4	75.3
C-6	27.1	27.0	26.9	26.6	26.8	26.8	26.6	26.6	26.9	37.0
C-7	21.6 <sup>b</sup>	21.4 <sup>a</sup>	21.2 <sup>a</sup>	20.9 <sup>a</sup>	21.6	20.6 <sup>a</sup>	20.2 <sup>a</sup>	24.0	21.9	18.1 <sup>b</sup>
C-8	41.9	41.8	41.8	41.6	41.8	41.5	41.2	36.7 <sup>a</sup>	41.3	42.2 <sup>c</sup>
C-9	35.8 <sup>a</sup>	35.8	35.9	35.8	36.1	36.2	36.8	45.1	32.6	40.2 <sup>c</sup>
C-10	35.8	35.8	35.6	35.4	35.8	35.5	35.4	36.2	35.5	55.8
C-11	21.7 <sup>b</sup>	21.9 <sup>a</sup>	21.3 <sup>a</sup>	21.4 <sup>a</sup>	21.6	21.2 <sup>a</sup>	21.3 <sup>a</sup>	21.4	30.0	22.8 <sup>b</sup>
C-12	40.4	41.2	41.0	40.9	40.3	31.3	40.6	37.7	74.8	40.2
C-13	50.3	50.4	50.7	50.5	50.3	49.5	52.6	54.2	56.4	50.1
C-14	85.6	85.2	84.1	83.8	85.6	86.1	85.7	146.3 <sup>c</sup>	85.8	85.3
C-15	33.0	42.6	39.5	39.3	33.0	31.3	38.8	108.3 <sup>d</sup>	33.0	32.2
C-16	27.3	72.8	75.0	74.7	27.3	24.8	133.8	135.8 <sup>d</sup>	27.9	27.5 <sup>a</sup>
C-17	51.5	58.8	56.8	56.6	51.5	48.9	161.2	158.0 <sup>c</sup>	46.1	51.4
C-18	16.1	16.9	16.1	16.1	16.0	18.5	16.6	20.1	9.4	16.2
C-19	23.9	23.9	23.9	23.8	23.9	24.0	24.1	24.0	23.8	195.7
C-20	177.1 <sup>c</sup>	171.8 <sup>b</sup>	171.5 <sup>b</sup>	171.5 <sup>b</sup>	177.1 <sup>a</sup>	173.6 <sup>b</sup>	172.8 <sup>b</sup>	173.5 <sup>b</sup>	177.1 <sup>a</sup>	177.2 <sup>d</sup>
C-21	74.5	76.7	76.8	76.5	74.7	74.8	72.6	72.1	74.6	74.8
C-22	117.4	119.6	121.3	121.1	117.4	116.6	111.7	119.5	117.0	117.8
C-23	176.3 <sup>c</sup>	175.3 <sup>b</sup>	175.8 <sup>b</sup>	175.4 <sup>b</sup>	176.3 <sup>a</sup>	175.8 <sup>b</sup>	176.3 <sup>b</sup>	176.8 <sup>b</sup>	176.3 <sup>a</sup>	176.6 <sup>d</sup>

OCOCH<sub>3</sub>, 21.3 ± 0.3; OCOCH<sub>3</sub>, 171.4 ± 0.4 ppm in III-VII.

a, b, c, d Values in any vertical column may be reversed.

involved in two 1,3-diaxial interactions, the C-6 axial hydrogen suffers steric compression only from the C-19 methyl group. The expected consequence of introduction of the C(14) : C(15) double bond in VIII and the C-12 $\beta$  hydroxyl group in IX differentiates the methine carbons, C-8, C-9, and C-5, the latter being unaffected. Differentiation of the signals of C-3, C-14, C-16, and C-23 depended on SFORD (1a) experiments and acetylation effects.

The  $^{13}\text{C}$  NMR data on strophanthidin (X) are in consonance with assignments of the signals for the other compounds presented here. The C-5 $\beta$  hydroxyl group in X unsymmetrically deshields the C-4 and C-6 methylene carbons owing to differences between axial and equatorial  $\beta$ -effects, respectively. The assignment of these signals, as well as that of the C-3 signal, was made from comparison with those in the spectra of some glycosides.<sup>†</sup>

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